



The science and practice of making feed enzyme decisions - the case for and against

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Enzyme Application to Feed

One of the most researched fields in poultry science

More than 2500 independent tests of feed enzymes in broilers (Rosen, 2010)

Grown to be a >\$550 million Industry that saves the global feed market ~ \$3 to 5 billion per year (Adeola & Cowieson, 2011).

Phytase A.niger C.braachi E.coli Citrobacter spp Buttiauxella spp





Xylanase ß-Glucanase Protease

Bacillus subtilis Bacillus licheniformis Other

Amylase Mannanase Galactosidase Glucoamylase Lipase



wiseGEEK

What drove the high penetration of feed enzymes we have today?

"Necessity is the mother of invention" William Horma, 1519



However, in spite of:

- Over >2500 research studies in enzymes by 2010
- Phytase application into >94% of broiler feed
- NSP/Carbohydrase /Protease in majority of broiler feed
- Every enzyme company having "scientific" matrix values for every conceivable enzyme



Not a lot of clarity on which enzymes are appropriate, factors causing variation in enzyme response, or additivity of enzyme matrix values in energy, let alone amino acid effects

Processes to select enzymes & combinations

1

{nowledge of substrates
 in feed ingredients



Match Enzyme Biochemistry to Substrates and Digestive Physiology *in-vitro& in-vivo* 3

In-Vivo Response



Performance (BW/FCR) Ileal Digestibility AMEn Gut health / Livability Increased Profitably

Key Decisions: Phytase

Which Phytase? What dose?

How much AvP / Ca²⁺ contribution?

Energy and Amino Acids from Phytase?

Table 1. Some examples of currently commercially available 3- and 6-phytases and their characteristics						
Type [†]	Protein origin	Expression	pH optima	Temperature optima (°C)	Trade name	
3	A. niger [*]	A. niger	2; 5–5.5	65	Natuphos®	
3	A. niger [*]	A. niger, non-recombinant	6.0	-	Allzyme [®] SSF	
3	A. niger [*]	Trichoderma reesei	2.5	-	Finase [®] P/L	
6	Escherichia coli*	Schizosaccharomyces pombe (ATCC 5233)	4.5	55	Phyzyme [®] XP	
6	Escherichia coli*	Pichia pastoris	4.5	-	Quantum®	
6	Escherichia coli	Trichoderma reesei	_	_	Quantum Blue®	
6	Escherichia coli [*]	Pichia pastoris	3.4, 5.0	58	OptiPhos [®]	
6	Peniophora lycii [*]	Aspergillus oryzae	4-4.5	50-55	Ronozyme [®]	
6	Citrobacter braakii	Aspergillus oryzae	-	-	Ronozyme Hiphos®	
6	Buttiauxella spp.	Trichoderma reesei	3.5-4.5#	60#	Axtra [®] PHY	
* Adapte † 3- or 6- #persona	*Adapted from Lei <i>et al.</i> ¹ with modifications; [†] 3- or 6-phytase; —, no information available; [#] personal communication (C Evans). Dersjant-Li et al, 2015					

Supplier Recommended Nutrient Contributions from Standard Dose of Phytase

	E.Coli 1	E.Coli 2	E.Coli 3	Citrobacter	E.Coli 4	Buttiauxella
Units/kg feed	500 FTU	500 OTU	500 FTU	1000 FYT	500 FTUQ	500 FTU
Digestible P%	0.11	0.11	-	0.117	-	0.134
"Available" P %	0.12	0.13	0.13	0.146	0.15	0.146
Calcium %	0.11	0.13	0.14	0.18	0.165	0.134
Phytase cost (\$/Feed Ton)	0.5	0.5	0.5	0.5	0.5	0.5
Phytase Cost / 0.12% AvP	0.50	0.46	0.46	0.41	0.40	0.41

Commercial values, 2014

In practice, decisions of phytase source and dose are frequently determined on phytase cost /0.10% or 0.12% AvP release Dose is usually < Max. profit from P replacement to risk



Methodology of how nutrient contributions from phytase were determined differs between commercially available phytases sources & affects decisions

Ileal vs. Tibia ash method; Adaptation time to test diets; Age broiler; Ca level & source Li et al.. 2014

	E.Coli 1	E.Coli 2	E.Coli 3	Citrobacter	E.Coli 4	Buttiauxella
FTU/kg feed	500 FTU	500 OTU	500 FTU	1000 FYT	500 FTU _Q	500 FTU
Digestible P %	0.11	0.11	-	0.117	-	0.134
Av.P %	0.12	0.13	0.13	0.146	0.15	0.146
Dig. P:AvP	0.92	0.85	-	0.80	-	0.92
Calcium %	0.11	0.13	0.14	0.18	0.165	0.134
Ratio Ca:AvP	0.92	1.00	1.08	1.23	1.10	0.92

Critical questions to ask to ensure you are comparing

in matrix

- What Research / Methodology was used to derive phosphorus (P) contribution?
- How does the P-system used compare to your Ingredient P matrix?
- What about Ca²⁺ matrix values? How were they determined?
- Is Phosphorus release in the matrix correlated with amino acid release / extra phosphoric effects of amino acids and energy?

Phytate not only affect phosphorus digestibility, but also amino acid digestibility, starch digestibility, endogenous losses, and live-performance



Higher phytate has also been shown to decrease live performance Woyengo et al., 2014

Animal type	Initial age (d)	PA ^z content in control diet (%)	PA content in PA diet (%)	Response criterion ^y	Decrease in performance due to PA (%)	Reference
Broiler	0	0.78	1.57	BWG	3	Liu et al. (2009)
Broiler	0	0.78	1.57	BWG	3	Liu et al. (2008a)
Broiler	0	0.78	1.57	BWG	7	Liu et al. (2008b)
Broiler	7	1.04	1.57	BWG	7	Cabahug et al. (1999)
Broiler	8	0.00	1.65	BWG	28	Onyango and Adeola (2009)

Phytase decisions on Source and Dose also need be based on phytate interactions with nutrients and understanding differences in biochemistry between phytase enzymes in the context of digestive physiology



- 1. Interactions of Phytate, Calcium, and Phytase Enzymes affects P contribution
- 2. Interactions of Phytate with Protein, Starch, and Na Anti-nutrient effects on live performance & drives ME& AA digestibility improvement from phytase
- 3. Differences in phytase enzyme pH optima and kinetics affect in-vivo results

Interactions of phytic acid with dietary nutrients are pH dependent

Mineral cations also chelate at low pHs if soluble (Tamin et al., 2003) Proteins and phytate acid also interact at higher pHs >6 in presence of Ca²⁺ Briggs (1959, Saio et al. (1967,1968)

Gizzard / Proventriculus

Duodenum / Ileum / Jejenum



Nelson et al., 1968; Maga, 1982; Angel et al., 2002, Selle et al., 2009, 2012; Walk et al., 2012

Interactions of Phytate and Ca²⁺

 Potential for phytate binding Ca²⁺ and other minerals increases with pH and dependent on Calcium source solubility Angel et al., 2002, Li et al., 2014





Affinity and binding strength of phytate esters with Ca²⁺ decreases IP6-IP3
 Luttrell (1993)

Phytase enzymes that can rapidly hydrolyze IP6-IP3 in Gizzard/ proventriculus will be less inhibited by Ca-phytate interactions

Phytic Acid Interactions with Protein

- Protein-Phytate complexes- form directly with phosphate group at low pH
- Tertiary bridges via Ca and basic residues in the protein , at pHs>6
- Protein-phytate formation proportional to the ratio of Phytate:Protein



Kies et al., 2006.

Yu et al.,2012 J. Anim Sci. 90:1824-32.

Protein-phytate complex formation is fundamental to phytate effects on protein/amino acid availability

Selle et al., 2012, Adeola&Cowieson, 2014

Only IP6 and to a lesser extent IP5 has the ability to aggregate with soluble proteins at a pH of 2.5



Phytase that can effectively degrade IP6-protein complexes rapidly at low pH will be more effective at

Large differences exist between phytase enzymes in optimum pH and enzyme kinetic properties



Published work on Citrobacter phytase shows this phytase seems to struggle degrading IP4 (DL-Ins(2,3,4,5) and IP3 esters.



Pontoppidan et al., 2012 in Arch.An.Nutrition, 66:6, 431-444

Differences in enzyme kinetics and pH optima of phytases result in very different phytate dephosphorylation patterns and phopsphate release during in-vitro simulation of digestion

Mendez et al., **2015**, J.Agric Chem.



Enzymatic phytate dephosphorylation of wheat during in vitro simulation of poultry digestive tract in a high buffer system

Degradation of protein-phytate complexes or Na-phytate by phytase



All values expressed relative to release of iP by Buttiauxella phytase on sodium phytate substrate as 100%

Differences in In-vitro phytase chemistry , IP6 hydrolysis rate & protein-phytate degradation need to be supported by repeatable in-vivo responses



Plumstead et al., 2012

What about other Enzymes other than Phytase?

Knowledge of substrates in feed ingredients



Match Enzyme Biochemistry to Substrates and Digestive Physiology *in-vitro& in-vivo* 3

In-Vivo Response



Performance (BW/FCR) Ileal Digestibility AMEn Gut health / Livability Increased Profitably

Xylanase / B-glucanase is normally the first enzyme considered for wheat/barley based diets... and corn

Substrates	Examples	Main anti-nutritive effects		
Soluble, non-viscous,	Stachyose	Increased activity of intestinal flora		
α-galactosides	Raffinose	Increased osmolarity and reduced DM of digesta		
		Increased digesta viscosity		
Soluble viscous NSPs		Increased mean retention time of digesta		
	Arabinoxylans and β-glucans (low molecular weight)	Reduced absorption rate of nutrients		
		Increased activity of intestinal flora		
Insoluble, non-viscous, NSPs	Arabinoxylans and β-glucans (high molecular weight)	Reduced accessibility of nutrients (e.g.		
	Cellulose	physical entrapment of starch granules		
	Starch, Resistant Starch	Reduced ME value of ingredients		
Starch	Varyable Amylose:Amylopectin, Starch- protein complexes	Increased substrate for gut microbiota		
	Variable digestibility of protein / AA,	Reduced ME + AA value of ingredients		
Protein	especially in poorer quality ingredients	Increased substrate for gut microbiota		
Dhutata	Variable amounts in feed, antinutritive	Reduced Ca, P, ME, AA digestibility		
Ρηγιαιε	effects other than Phosphorus	Interactions with gut microbiota		

Arabinoxylan and beta-glucan in some feed ingredients (% dry matter)



To be effective in reducing Viscocity of soluble NSPs, a Xylanase needs to be able to hydrolyze both Insoluble and soluble arabinoxylan fractions Choct et al., 2004; Adeola and Cowieson, 2014)

NSP database Source: Choct (2006); Danisco Non Starch Polysaccharide (NSP) database (2012)

Arabinoxylans from cereals are structurally complex and differ between feed ingredients in structure of Arabinose side chains and Diferulic bridges



Although Xylanase targets ArabinoXylan substrate... there seem to be source-dependent differences in response



DuPont Laboratory, 2010 unpublished

In-Vivo support of xylanase being effective in both corn and wheat-based diets is required

			R	etention, % of intake			
$\frac{1}{\alpha}$ Main	n effect	NDF retention%	ADF retention%	AMEn Kcal/kg	NDF ¹	ADF ¹	AME _n , kcal/kg
Cont	trol	27.9 ^b	9.57 ^b	2995 ^b	31.7 36.2 24.0	11.9 19.4 7.23	3,005 3,061 2,985
+ Xy	lanase	32.3ª	16.6ª	3059ª	28.4 1.022	13.9 2.29	3,057 16.18
Prob	ability				.9ª	$\frac{15.7^{\rm a}}{10.6^{\rm b}}$	$3,033 \\ 3,026$
Diet		<0.01	0.04	<0.001		1.62	11.44
Xyla	nase	<0.01	0.01	< 0.001	27.9 ^b 32.3 ^a 0.72	9.57 ⁶ 16.6 ⁸	2,995 ⁶ 3,059 ^a
^I Diet x Xy	lanase	0.95	0.86	0.63	<0.01 <0.01 0.95	0.04 0.01 0.86	0.45 <0.01 0.63

Kiarie, Romero, and Ravindran, 2014

Dose response trials to Xylanase in 42-d Broilers are sometimes frustrating with variable responses that are hard to predict

Simple corn-soy diets, single Xylanase dose

Plumstead, 2009, unpublished

Bio-efficacy of exogenous Xylanase and other enzymes may be affected by complex interactions between substrates in the feed ingredient and with the gut biome

Consequently, reported performance responses have been variable

In addition to NSP's do we also need to consider other substrates when selecting enzymes for corn/soy –based diets?

Substrates	Examples		Main anti-nutritive effects		
Soluble, non-viscous,	Stachyose		Increased activity of intestinal flora		
α-galactosides	Raffinose In addition		on to ''NSP's,		
Soluble, viscous, NSPs	Arabinoxylans and β- molecular wei	Undiges Protein largest a "Substra mixed c	account for the amount of undigested ate" available in orn/soy-based diets		
Insoluble, non-viscous, NSPs	Arabinoxylans and β-glucans (high molecular weight) Cellulose		Reduced accessibility of nutrients (e.g. physical entrapment of starch granules)		
Starch, Resistant		Starch	Reduced ME value of ingredients		
Starch	Varyable Amylose:Amylo protein comple	pectin, Starch- exes	Increased substrate for gut microbiota		
Protein	Variable digestibility of protein / A especially in poorer quality ingredie		Variable digestibility of protein / AA, especially in poorer quality ingredients		Reduced ME + AA value of ingredients Increased substrate for gut microbiota
Phytate	effects other than Ph	, antinutritive losphorus	Interactions with gut microbiota		

Corn morphology is important to degree of starch-protein binding, degree of starch digestion and responsiveness to enzymes

Vitreous Endosperm

Floury Endosperm

Starch Prolamin granule Zein Protein matrix

Scanning electron microscopy of starch granules in corn: A) starch granules heavily imbedded in prolamin-protein matrix, B) starch granules in opaque corn endosperm with less extensive encapsulation by prolamin-proteins (Gibbon et. al., 2003).

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Assessing variation in Corn protein Composition

- The amount of Prolamin-Zein protein can be quantified analytically Hamaker et al., 1995 – Cereal Chemistry
- % prolamin of total protein is affected by growing conditions, maturity, cultivar, and drying conditions of corn

126 corn samples, 8 different countries over 2 years

DuPont, Internal data

In vitro effects of graded α-amylase dose on corn with high (80) of low (20) Proma values

DuPont, Internal data

Ileal starch digestibility in broilers: 15 digestibility trials with XA (Xylanase+Amylase) or XA+Protease (XA+P)

Decisions on Protease in Broiler diets?

> Protease effects in feed

1. Hydrolysis of dietary protein and increased protein solubility

(Caine et al., 1998)

- 2. Disruption of protein-starch interactions in corn (Mc Allister et al., 1993; Belles et al., 2000)
- 3. Disrupt Fibre-protein interactions

Colombatto and Beauchemin, 2009

4. Potential gut health benefits of reducing fermentation of undigested protein in ceca/colon

Other benefits of Protease: Fibre Digestion by Xylanase!

- Serine protease tested in digestion of alfalfa in rumen batch model
- Protease increased in vitro disappearance of DM, NDF, hemicellulose

From Colombatto and Beauchemin, 2009 Figure 4. Scanning electron microscopy images of alfalfa hay samples, untreated (a) or enzyme-treated (b; Protex 6L, Genencor Int., Rochester, NY) at 0 h, or untreated (c) or enzyme treated (d) at 18 h postincubation with runnial fluid in vitro.

Effect of Xylanase Source and Protease dose on Soluble Pensosan release from Corn DDGs

Pedersen et al.Unpublished

Bio-efficacy of exogenous enzymes is not only related to the primary biochemical target of enzymes

Phytase	P and Ca digestibilityA.A., fat digestibility
Xylanases, B-glucanse	Fibre disappearanceA.A., fat, starch digestibility
Amylases	 Starch, A.A. digestibility
Proteases	A.A. / Protein digestibilityFibre digestibility?
Mannanase	Galactomannan degradationReduction in Innate Immune response

Hsiao et al., 2006; Romero & Plumstead 2014

Decisions on enzyme addition to feed with phytase

- 1. General consensus that enzyme effects are **NOT additive** with responses ranging from antagonistic to synergystic
- Enzyme Response is based on law of diminishing returns. As phytase is included in >>94% of Broiler feed ...
 ^{Cowieson et al., 2012}
 ...any other additive need to demonstrate value on TOP of
 phytase, and each other.

Combining enzyme activities needs to make sense in terms of substrates and be quantifiable in biological trials

Cowieson and Adeola, 2014

Overall objective: address unknown variation by improving mean and consistency of live performance

XA+P enzyme applied to 26 different corn samples fed to broilers

Application of enzymes in poultry diets: Simplifying complexity

- Enzyme responses are dependent on dietary Substrates they target, which we need to understand better.
- Value of Phytase is far greater than improvements in phosphorus availability and negative effects of phytate on nutrient utilization and performance need to be considered in decision making process.
- Enzyme effects are sub-additive, based on a law of diminishing return
- Value of carbohydrases and other enzymes must be determined on top of phytase
- Some assessment of feed ingredient quality is required to explain variation in enzyme responses

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